

6. Add extraction-loading buffer to the pellet. Use 15  $\mu$ l of the buffer per each 20  $\mu$ l of Afyon resin initially added to the protein solution in step 2.
7. Vortex the mixture for 1 minute.
8. Transfer the mixture of the buffer with Afyon resin into the spin filter insert of the filtration device included in the kit.
9. Centrifuge the spin filtration device at maximum speed (14,000 rpm) for 1 min to filter out the resin.
10. Discard the filter insert with used resin.
11. The resulting collected filtrate is ready to load on an SDS-gel.

### For More Information



including a detailed manual, visit [www.advansta.com/Afyon](http://www.advansta.com/Afyon) or go directly to the web-page by scanning the QR-code with your favorite bar-code scanner app on your smart phone.

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*Afyon*<sup>TM</sup>

**Fast, easy concentration and preparation of protein samples for SDS-PAGE**

### For Catalog Numbers

**K-02101-010**

Afyon SDS-PAGE Sample Preparation Kit, 10 samples

**K-02101-025**

Afyon SDS-PAGE Sample Preparation Kit, 25 samples



## Storage Information

Afyon resin is supplied as a 25% slurry in water. For long term storage, keep the tube containing the resin upright in a rack at 4° C. Do not freeze, boil, or autoclave the resin. Protect the resin from long exposure to bright light, and do not allow the resin to dry. Afyon extraction-loading buffer may be stored at room temperature or below. If it was stored refrigerated, bring it to room temperature before use and make sure any precipitate that could form during storage is completely dissolved.

## Warnings and Precautions

- Afyon resin is for research use only.
- Always wear gloves when handling reagents.
- Refer to MSDS for additional safety information.
- The product is guaranteed to be free of manufacturer defects, and to function as described when the provided protocol is followed by properly trained personnel.

## Short Protocol

### Step

1. Vigorously vortex the tube of Afyon resin to resuspend it to homogeneity.
2. Add 20 µl of homogeneous Afyon resin to the protein solution. This amount will be sufficient to obtain a protein sample for one lane of a typical mini-gel (~5 µg of total cellular protein). If more than one identical protein lane is required, increase the volume of Afyon resin accordingly. If your protein solution contains high concentrations of salts, detergents or chaotropic agents, dilute the solution first to acceptable levels. Refer to the compatibility chart or to our web-site for details.
3. Vortex the mixture for 30 seconds if the dilution factor for the resin is 1:100 or less (your sample fits in a standard 1.5ml or 2ml micro-centrifuge tube). For higher dilution factors (sample volumes larger than 2ml) increase vortexing time (up to 5 minutes for 50ml samples).
4. Centrifuge the tube to pellet the resin.
5. Remove the supernatant. If using micro-centrifuge tubes, remove the supernatant as completely as possible, making sure not to disturb the blue pellet of the resin. If the total volume is too large for a micro-centrifuge tube, remove the supernatant in two steps, as follows. 1) Remove most of the supernatant and leave 0.5ml to 1ml of it in the tube. 2) Resuspend the pellet in this remaining solution and transfer the mixture into a micro-centrifuge tube. Spin down again and remove the supernatant as completely as possible.