Xpert Taq^{PLUS} Hotstart Mastermix (2X)

#GE19.0001 (1ml) | GE19.5001 (5x 1ml)

(FOR RESEARCH ONLY)

Product: Xpert Taq^{PLUS} Hotstart DNA polymerase is an enhanced DNA polymerase with superior PCR performance when it comes to the amplification of long templates (up to 30kb). The buffer composition has been optimized via high-throughput screening and the enzyme is particularly efficient for the amplification of difficult templates, such as mammalian genomic DNA, and GC-rich or AT-rich templates. As a result, Xpert Taq^{PLUS} DNA Polymerase is this the ideal choice for consistent results in complex PCR amplifications. Upon completion of PCR, the reaction is ready for direct loading onto an agarose gel without the need of adding loading buffer.

Applications: Very difficult templates, Long PCR

Contents: Xpert Taq^{PLUS} Hotstart DNA Polymerase is supplied as a convenient 2x mastermix, including an inert red tracking dye for electrophoresis, containing all required components for fast PCR, except specific primers. Final concentration of $MgCl_2$ will be 3mM. One ml is suitable for 80 reactions of 25µl (or 100 reactions of 20µl).

Component	GE29.0001	GE29.5001
Xpert Taq^{PLUS} Hotstart Mastermix (2X) with dye	1 ml	5x 1ml

QC: Functionally tested in PCR. Absence of endonucleases, exonucleases, and ribonucleases was confirmed by appropriate assays

Properties:
Amplicon size: up to 30kb
Extension rate: 4kb/min (for targets up to 5kb) and 1kb/min for larger targets
Hotstart: Yes
A-overhang: Yes

Storage: -20°C and protected from light for at least 1 year. No loss of performance is detected after 20 freeze/thaw cycles.

Prior to use

Optimal PCR cycling conditions (incubation times and temperatures) depend on DNA target (GC-content, size, quantity, purity, etc.) and specific primers and need to be determined case by case. Xpert Taq^{PLUS} Hotstart Mastermix (2X) with dye includes dNTPs and has already been optimized with respect to the $MgCl_2$ concentration and other components to maximize success rates. It is not recommended to add additional $MgCl_2$ or other PCR enhancers. We suggest to start with the following basic protocol and subsequently optimize annealing temperature, incubation times and cycling number.

Basic Protocol

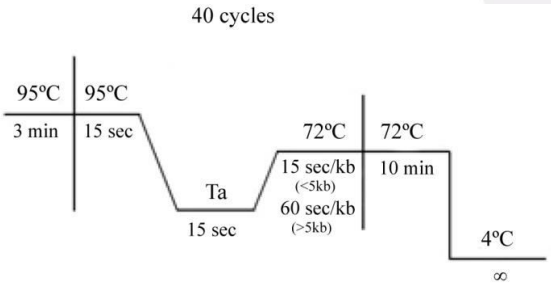
1. Mix for each PCR reaction, starting with the greatest volume (usually water):

Component	Volume (25µl)	Final Conc.
Xpert Taq^{PLUS} Hotstart Mastermix (2X) with dye	12.5 µl	1X
Forward primer (5 pmol/µl)	2 µl	0.4 µM
Reverse primer (5 pmol/µl)	2 µl	0.4 µM
Template DNA*	0.25 – 10 µl	1-250 ng*
PCR–grade water	up to 25 µl	

**) In case of cDNA <50ng and in case of gDNA <250ng (total amount).*

In order to minimize risk of contamination, reagent loss and improve pipetting accuracy, we recommend to prepare a mastermix for multiple samples (N), always including a negative control for the detection of possible contaminants, by mixing all components (N+1), except template DNA, dividing the mixture equally into each tube and then add template DNA or PCR grade water in case of the control to the individual PCR tubes.

2. Set-up initial PCR amplification as follows:



After an initial cycle of 3 min at 95°C (enzyme activation and denaturation of template DNA), cycle 40 times for 15 seconds at 95°C, 15 seconds at Ta, and either 15 seconds per kb of target DNA for amplicons up to 5kb or 60 seconds per kb of targets for amplicons between 5kb and 30kb) at 72°C for extension. Set the annealing temperature (Ta) as the melting temperature (Tm) of the primer with the lowest Tm. After amplification, include a final extension step of 10 min at 72°C to ensure that all amplicons are fully extended and include 3´-A-overhang. Analyze PCR products by DNA Agarose gel electrophoresis. Samples can be loaded directly onto an agarose gel without the need of adding a loading buffer. Using a 1% agarose gel, the inert red tracking dye co-migrates with DNA of approximately 600bp and using a 2% with DNA of approximately 350bp.

Optimization

Annealing Temperature (Ta) and Primers

Optimizing the annealing temperature is crucial, as a too low temperature might result in non-specific amplification whereas a too high temperature results in no amplification. The melting temperature (Tm) is defined as the temperature in which 50% of the primer and its complementary sequence of the target DNA are present as duplex DNA. By increasing the temperature above the melting temperature, this percentage decreases, however, primers will still anneal (up to a certain point) and initiate extension. PCR can therefore be performed at temperatures of several degrees higher than Tm and it is therefore recommended to optimize the Ta by performing a temperature gradient (e.g., starting at the lowest Tm or a few degrees below and increasing with 2°C increments). Ideally, primers have melting temperatures of approximately 60°C and final concentration should be between 0.2 and 0.6µM (each).

Incubation times and number of cycles.

Denaturation and annealing steps may require less time depending on the thermocycler apparatus (ramp rate), reaction volume and PCR tube (varies with the efficiency of heat-transfer). It might be worthwhile to optimize (reduce) times to as low as 10 seconds for both denaturation and annealing steps, which will greatly reduce overall PCR time. It might be worthwhile to reduce number of cycles from 40 to 25-30, depending on the success of amplification.