# Coomassie Brilliant Blue <br> (FOR RESEARCH ONLY) 


#### Abstract

Product: Coomassie Brilliant Blue G-250 (CBB) is widely used for visualizing proteins after electrophoresis.


Applications: In-gel staining of proteins. Suitable for acrylamide and agarose gels.
Quantity: \#GS22.1000 contains 1L of a ready-to-use Coomassie Brilliant Blue Solution
Appearance: Dark Blue solution
Storage: Store at room temperature, protected from light, for at least 2 years

## Usage:

## Standard Gel Staining Protocol

1. After electrophoresis, place the gel in a suitable container and wash once or twice ( $10-30$ seconds) with deionized water.
2. Gels may be prefixed in $45 \%$ Methanol, $10 \%$ Acetic Acid, $45 \%$ deionized water for $30-60$ minutes (alternatively overnight).
3. Rinse the gel several times with deionized water to completely remove fixing solution.
4. Stain the gel with Coomassie Brilliant Blue staining solution and incubate with gentle agitation for 2 to 4 hours (until gel is uniform blue and no longer clearly visible in the staining solution).
5. Destain in $5 \%$ Methanol, $7.5 \%$ Acetic Acid, $87.5 \%$ deionized water until background is clear (4-24 hours). Strongest bands should appear after 1-2 hours. If background is completely clear, this should allow for the detection of as little as $0.1 \mu \mathrm{~g} / \mathrm{band}$.
6. Once bands are visually strong enough, discard the destaining solution.
7. Gel is ready for imaging and analysis or storage (e.g., in $7 \%$ acetic acid).
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[^0]:    * Fixing/Fixation denatures and precipitates proteins within the gel matrix and prevents the diffusion of proteins resulting in sharper bands. Fixation also removes buffer components, most importantly SDS that may interfere with staining. Small and extremely soluble proteins may require stronger fixing solutions, e.g., based on TCA.

